

## Valence Tautomerism and Coordinative Lability in Copper(II)–Imidazolyl–Semiquinonate Anion Radical Models for the Cu<sub>B</sub> Center in Cytochrome *c* Oxidases

David G. Lonnon, Sang Tae Lee, and Stephen B. Colbran\*

School of Chemistry, University of New South Wales, Sydney, NSW 2052, Australia

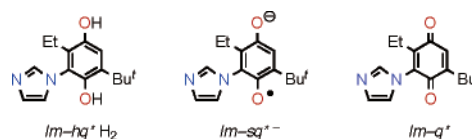
Received January 7, 2007; E-mail: s.colbran@unsw.edu.au

We report that internal electron transfer (valence tautomerism) within a Cu<sup>II</sup>–imidazolyl–phenolate anion system can lead to a substitutionally labile Cu<sup>I</sup>–imidazolyl–phenoxy radical system. These results inform about the potential reactivity of a Cu<sub>B</sub><sup>II</sup>–histidine–tyrosinate intermediate in the catalytic center of cytochrome *c* oxidase (CcO).

In the final steps of the aerobic respiratory transfer chain, dioxygen is reduced to water within the heme a<sub>3</sub>••Cu<sub>B</sub> catalytic center (Figure 1) of CcO.<sup>1</sup> Free energy is conserved by coupled translocation of protons through CcO and across an inner mitochondrial or bacterial membrane, thereby maintaining the proton electrochemical gradient that drives ATP synthesis. The scission of dioxygen occurs in the very first step of the CcO catalytic cycle and requires four electrons donated under turnover conditions by Fe<sub>a3</sub><sup>II</sup>, Cu<sub>B</sub><sup>I</sup>, and Tyr244, affording the so-called P<sub>M</sub> intermediate comprising ferryl Fe<sub>a3</sub><sup>IV</sup>=O, cupric Cu<sub>B</sub><sup>II</sup>–OH, and, more controversially, tyrosyl radical TyrO• centers.<sup>2,3</sup> Microsecond freeze hyperquenching experiments suggest the P<sub>M</sub> state forms upon dioxygen scission regardless of the initial oxidation level of the enzyme.<sup>4</sup> In the next (and the first proton-pumping) step of the catalytic cycle, the tyrosyl radical is the electron acceptor, leading to the possibly obligatory intermediacy of a tyrosinate Cu<sub>B</sub><sup>II</sup>–His–TyrO<sup>−</sup> species.

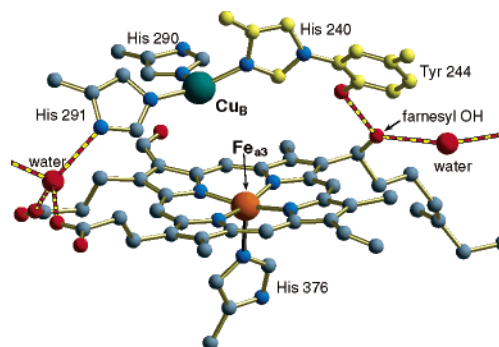
Mimics for Cu<sub>B</sub><sup>II</sup>–His–TyrO• and Cu<sub>B</sub><sup>II</sup>–His–TyrO<sup>−</sup> centers would provide valuable physicochemical information for comparison with biophysical data.<sup>3,6,7</sup> One major problem with the models thus far investigated is that the imidazolyl–phenoxy radicals derived from them are only transiently stable.<sup>3,6,7</sup> In contrast, semiquinone anions (i.e., phenoxy–phenoxide radicals), which may be generated by one-electron reduction of quinones or, conversely, by one-electron oxidation of hydroquinone dianions, are stable, long-lived species under anaerobic, basic conditions.<sup>8</sup> Hence, to avoid the limitations due to decomposition reactions of imidazolyl–phenoxy radicals, we have targeted copper complexes of the semiquinone radical, *Im*–*sq*<sup>•−</sup>. In this work, the hydroquinone/semiquinone group acts as a surrogate for the phenol/phenoxy radical side chain of the cross-linked tyrosine in CcO.

Akin to an ordinary hydroquinone, *Im*–*hq*<sup>•−</sup>H<sub>2</sub> readily undergoes two-electron oxidation to the corresponding quinone *Im*–*q*<sup>•</sup> (e.g., with phenylidodisacetate, Ag<sub>2</sub>O, or 2,3-dichloro-5,6-dicyano-1,4-benzoquinone). One-electron oxidation of the hydroquinone dianion, *Im*–*hq*<sup>•2−</sup> (from *Im*–*hq*<sup>•−</sup>H<sub>2</sub> and lithium diisopropylamide (LDA), 2.0 equiv) or KOBu<sup>t</sup> (2.0 equiv)), and one-electron reduction of *Im*–*q*<sup>•</sup> are chemically reversible processes and afford the semiquinone radical anion, *Im*–*sq*<sup>•−</sup>.<sup>9</sup> The distinctive UV–vis and EPR spectra<sup>10</sup> of *Im*–*sq*<sup>•−</sup> epitomize those of a semiquinone or phenoxy radical;<sup>11</sup> in the EPR spectrum, the *N*-superhyperfine coupling is notably small (~0.5 G), indicative for only slight inter-ring spin delocalization and in line with theoretical predictions for the histidine–tyrosyl radical in CcO.<sup>3,5</sup>

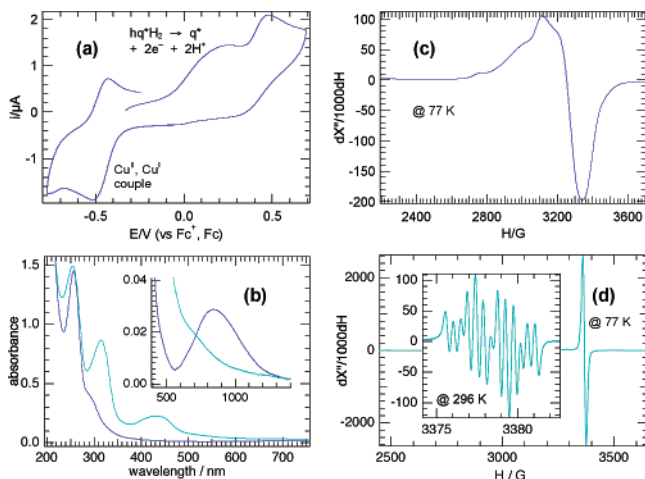


The Cu<sup>II</sup>–imidazolyl–hydroquinone complex [(tpa)Cu(*Im*–*hq*<sup>•−</sup>H<sub>2</sub>)](OTs)<sub>2</sub> (**1**; tpa = tris(2-pyridylmethyl)amine, OTs<sup>−</sup> = *p*-toluenesulfonate anion) was prepared by addition of *Im*–*hq*<sup>•−</sup>H<sub>2</sub> to [(tpa)Cu(MeCN)](OTs)<sub>2</sub> in anhydrous methanol/ether.<sup>9</sup> Separate deprotonations of **1** were carried out under anaerobic, anhydrous conditions in two solvents, acetonitrile or tetrahydrofuran, and using two different bases, KOBu<sup>t</sup> or LDA. Addition of 2 equiv of base results in bleaching of the *d*–*d* bands for **1** (λ<sub>max</sub> 865 nm) and the appearance of the intense, characteristic bands of *Im*–*sq*<sup>•−</sup> at 320 and 435 nm (Figure 2 and Supporting Information). In the EPR spectrum, the axial signal of **1** in MeCN is replaced by a sharp signal at *g* = 2.0051 for an organic radical. At room temperature, the EPR spectrum is identical to that for free *Im*–*sq*<sup>•−</sup> radical (see above). The estimated conversion to the free *Im*–*sq*<sup>•−</sup> radical anion in these experiments is 70–80%.<sup>9</sup> The <sup>1</sup>H NMR spectrum after double deprotonation of **1** shows sharp peaks for the Cu<sup>I</sup> complex, [Cu(tpa)(L)]<sup>+</sup> (L = solvent, HNPr<sub>2</sub> from LDA or HOBU<sup>t</sup> from KOBu<sup>t</sup>),<sup>12</sup> the residual protio-solvent, the base, and no others.<sup>9</sup> Entirely analogous behavior is observed in THF solution.<sup>9</sup> The copper-containing species in acetonitrile solution were also characterized by ESI-FT-ICR mass spectroscopy: before and after adding KOBu<sup>t</sup> (2.0 equiv) to **1**, the prominent ions were [(tpa)Cu(*Im*–*hq*<sup>•−</sup>H<sub>2</sub>)]<sup>2+</sup> (*m/z* 306.61752 (major isotopomer); calcd *m/z* 306.61761) and [(tpa)Cu]<sup>+</sup> (*m/z* 353.08192 (major isotopomer); calcd *m/z* 353.08094), respectively.

In toto, these results provide unequivocal evidence for formation of the free *Im*–*sq*<sup>•−</sup> radical and [Cu(tpa)(L)]<sup>+</sup> upon double deprotonation of **1**. The probable mechanism involves an internal electron transfer (valence tautomerism) producing a coordinatively

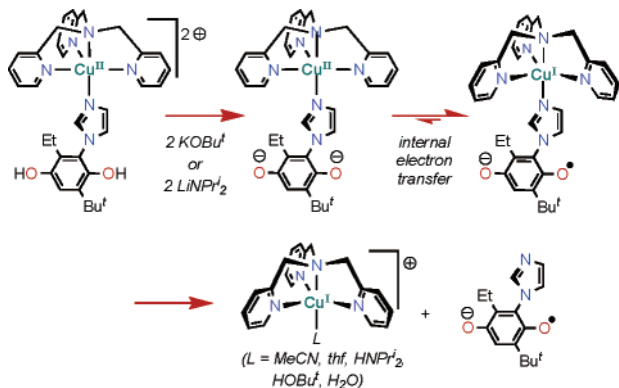


**Figure 1.** View of the binuclear heme a<sub>3</sub>••Cu<sub>B</sub> catalytic center of cytochrome *c* oxidase (coordinates and residue numbering for oxidized bovine enzyme, pdb accession no. 1V54, ref 5).



**Figure 2.** (a) Cyclic voltammogram of  $[(\text{tpa})\text{Cu}^{\text{II}}(\text{Im}-\text{hq}^*\text{H}_2)](\text{OTs})_2$ , **1** (2 mM) in MeCN–0.1 M  $[\text{Bu}_4\text{N}][\text{PF}_6]$ . (b) UV–vis–NIR (at 296 K) and (c, d) X-band EPR spectra for a solution of **1** (1.2 mM) in MeCN before (blue, –) and after (cyan, –) the addition of base ( $\text{KOBu}^t$ , 2.0 equiv). Simulation<sup>9</sup> of the 296 K EPR spectrum gives  $g = 2.0051$ ,  $a_{\text{H}}/G = 1.805$  (1 H) and 1.375 (2 H);  $a_{\text{N}}/G = 0.490$  (1 N).

### Scheme 1



labile copper(I) intermediate  $[(\text{tpa})\text{Cu}^{\text{I}}(\text{Im}-\text{sq}^{*-})]$  (Scheme 1). Consistent with this interpretation, cyclic voltammetry<sup>13</sup> reveals the  $\text{Cu}^{\text{II}}$ ,  $\text{Cu}^{\text{I}}$  couple for **1** is  $-0.47$  V, whereas the  $\text{Im}-\text{hq}^{*2-}$ ,  $\text{Im}-\text{sq}^{*-}$  and  $\text{Im}-\text{sq}^{*-}$ ,  $\text{Im}-\text{q}^*$  couples for uncomplexed  $\text{Im}-\text{hq}^{*2-}$  are  $\sim -1.5$  and  $-0.94$  V,<sup>9</sup> respectively. The  $\text{Im}-\text{hq}^{*2-}$ ,  $\text{Im}-\text{sq}^{*-}$  couple should be only slightly perturbed by coordination of the imidazole to a  $\text{Cu}^{\text{II}}$  center,<sup>14</sup> thus an internal electron transfer upon double deprotonation of **1** is expected. A phenoxyl radical, phenolate anion couple will always be higher than a closely related quinone, semiquinone couple; thus it is possible that  $\text{Cu}^{\text{II}}$ ,  $\text{Cu}^{\text{I}}$  and phenoxyl radical, phenolate anion couples become closely matched, for example, the phenoxyl radical, phenolate anion couple of the 4-acetoxy derivative of  $\text{Im}-\text{hq}^*\text{H}_2$  is  $-0.25$  V,  $\sim 700$  mV higher than the  $\text{Im}-\text{q}^*$ ,  $\text{Im}-\text{sq}^{*-}$  couple and higher than the  $\text{Cu}^{\text{II}}$ ,  $\text{Cu}^{\text{I}}$  couple in **1**.<sup>15</sup>

These results highlight the need to consider valence tautomerism and its possible effects in CcO. EXAFS studies reveal that one of the three histidine ligands to the  $\text{Cu}_{\text{B}}$  ion is more weakly bound than the other two and is coordinatively labile in the  $\text{Cu}_{\text{B}}^{\text{I}}$  state.<sup>16</sup> High-level theoretical calculations suggest that the  $\text{Cu}_{\text{B}}^{\text{II}}$  and the histidine–tyrosyl radical centers in the  $\text{P}_{\text{M}}$  intermediate have close to identical reduction potentials, that is, which center is the electron

acceptor is finely balanced<sup>17</sup> (although a presumption that the tyrosyl radical is the electron acceptor pervades the literature<sup>1–7,14–17</sup>). Thus, if a tyrosinate  $\text{Cu}_{\text{B}}^{\text{II}}-\text{His}-\text{TyrO}^-$  center occurs *anywhere* in the catalytic cycle—for example, if the reduction of the tyrosyl radical in the  $\text{P}_{\text{M}}$  state is not strictly concerted with the transfer of a proton to the tyrosyl oxygen atom thereby affording the tyrosinate anion<sup>1–7,14–17</sup>—valence tautomerism may lead to a (transient)  $\text{Cu}^{\text{I}}-\text{His}-\text{TyrO}^\bullet$  center. Dissociation of the labile histidine ligand from the thus formed  $\text{Cu}_{\text{B}}^{\text{I}}$  center could have important ramifications such as the redox-linked opening of a pathway during turnover for egress of pumped protons and/or product waters from the  $\text{Fe}_{\text{a}3}\cdots\text{Cu}_{\text{B}}$  catalytic center.

**Acknowledgment.** We are grateful for support from the Australian Research Council (Grant No. DPO557462).

**Supporting Information Available:** Details of preparations and measurements plus additional electrochemical, NMR, EPR, UV–vis–NIR spectra and data. This material is available free of charge via the Internet at <http://pubs.acs.org>.

### References

- (1) (a) Brzezinski, P. *Trends Biochem.* **2004**, *29*, 380–387. (b) Gennis, R. B. *Front. Biosci.* **2004**, *9*, 581–591. (c) Wikström, M. *Biochim. Biophys. Acta* **2004**, *1655*, 241–247.
- (2) (a) Proshlyakov, D. A.; Pressler, M. A.; Babcock, G. T. *Proc. Natl. Acad. Sci. U.S.A.* **1998**, *95*, 8020. (b) Proshlyakov, D. A.; Pressler, M. A.; DeMaso, C.; Leykam, J. F.; DeWitt, D. L.; Babcock, G. T. *Science* **2000**, *290*, 1588–1591. (c) Iwaki, M.; Puustinen, A.; Wikstrom, M.; Rich, P. R. *Biochemistry* **2004**, *43*, 14370–14378. (d) Iwaki, M.; Puustinen, A.; Wikström, M.; Rich, P. R. *Biochemistry* **2006**, *45*, 10873.
- (3) Barry, B. A.; Einarsson, O. *J. Phys. Chem. B* **2005**, *109*, 6972–6981.
- (4) (a) Cherepanov, A. V.; de Vries, S. *Biochim. Biophys. Acta* **2004**, *1656*, 1–31. (b) Wiertz, F. G. M.; Richter, O. M.; Cherepanov, A. V.; MacMillan, F.; Ludwig, B.; de Vries, S. *FEBS Lett.* **2004**, *575*, 127–130. (c) Wiertz, F. G. M.; de Vries, S. *Biochem. Soc. Trans.* **2006**, *34*, 136–138.
- (5) Tsukihara, T.; Shimokata, K.; Katayama, Y.; Shimada, H.; Muramoto, K.; Aoyama, H.; Mochizuki, M.; Shinzawa-Itoh, K.; Yamashita, E.; Yao, M.; Ishimura, Y. *Proc. Natl. Acad. Sci. U.S.A.* **2003**, *100*, 15304–15309.
- (6) Kim, E.; Chufan, E. E.; Kamaraj, K.; Karlin, K. D. *Chem. Rev.* **2004**, *104*, 1077–1133.
- (7) (a) McCauley, K. M.; Vrtis, J. M.; Dupont, J.; Van der Donk, W. A. *J. Am. Chem. Soc.* **2000**, *122*, 2403–2404. (b) Cappuccio, J. A.; Ayala, I.; Elliott, G. I.; Szundi, I.; Lewis, J.; Konopelski, J. P.; Barry, B. A.; Einarsson, O. *J. Am. Chem. Soc.* **2002**, *124*, 1750–1760. (c) Kim, S. H.; Aznar, C.; Brynda, M.; Silks, L. A. P.; Michalczuk, R.; Unkefer, C. J.; Woodruff, W. H.; Britt, R. D. *J. Am. Chem. Soc.* **2004**, *126*, 2328–2338. (d) Nagano, Y.; Liu, J. G.; Naruta, Y.; Ikoma, T.; Tero-Kubota, S.; Kitagawa, T. *J. Am. Chem. Soc.* **2006**, *128*, 14560–14570.
- (8) *The Chemistry of the Quinonoid Compounds*; Rappoport, Z., Patai, S., Eds.; John Wiley & Sons: New York, 1988; Vols. 1 and 2.
- (9) See the Supporting Information.
- (10) Data for  $\text{Im}-\text{sq}^{*-}$  generated by electrochemical oxidation of  $\text{Im}-\text{hq}^*\text{H}_2$  + LDA (2.0 equiv): UV–vis (THF),  $\lambda_{\text{max}}/\text{nm}$  ( $\epsilon_{\text{max}}/\text{M}^{-1}\text{cm}^{-1}$ ) 326 ( $14.3 \times 10^3$ ), 400 sh, 420 ( $7.1 \times 10^3$ ), 430 sh; X-band EPR, 296 K (MeCN):  $g_{\text{iso}} = 2.0051$ ,  $a_{\text{H}}/G = 1.81$  (1 H), 1.38 (2 H),  $a_{\text{N}}/G = 0.49$ .
- (11) Chaudhuri, P.; Wieghardt, K. *Prog. Inorg. Chem.* **2001**, *50*, 151–216.
- (12) Tyeklar, Z.; Jacobson, R. R.; Wei, N.; Murthy, N. N.; Zubieta, J.; Karlin, K. D. *J. Am. Chem. Soc.* **1993**, *115*, 2677–2689.
- (13) Potentials against the ferrocenium/ferrocene couple, which in acetonitrile is  $+0.630$  V vs NHE; see: Pavlishchuk, V. V.; Addison, A. W. *Inorg. Chim. Acta* **2000**, *298*, 97.
- (14) Colbran, S. B.; Paddon-Row, M. N. *J. Biol. Inorg. Chem.* **2003**, *8*, 855–865.
- (15) Lee, S. T.; Lonnon, D. G.; Craig, D. C.; Colbran, S. B. Submitted for publication.
- (16) (a) Osborne, J. P.; Cosper, N. J.; Stalhandske, C. M. V.; Scott, R. A.; Alben, J. O.; Gennis, R. B. *Biochemistry* **1999**, *38*, 4526–4532. (b) Ralle, M.; Verkhorvskaya, M. L.; Morgan, J. E.; Verkhorvsky, M. I.; Wikstrom, M.; Blackburn, N. J. *Biochemistry* **1999**, *38*, 7185–7194.
- (17) (a) Blomberg, M. R. A.; Siegbahn, P. E. M.; Babcock, G. T.; Wikstrom, M. *J. Am. Chem. Soc.* **2000**, *122*, 12848–12858. (b) Popovic, D. M.; Stuchebrukhov, A. A. *J. Am. Chem. Soc.* **2004**, *126*, 1858–1871. (c) Bu, Y. X.; Cukier, R. I. *J. Phys. Chem. B* **2005**, *109*, 22013–22026.

JA068972F